



Does gender selection works? An observational study

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Abstract

Sex selection is an option for couples who want to avoid passing a sex-linked genetic disease to their baby. For example, some females are carriers for over 350 X-linked diseases, even if the disease does not directly affect them. Hemophilia and Duchenne muscular dystrophy are examples of diseases that occur with a recessive gene on the X chromosome.

Some choose the sex of their baby because of a personal preference like "Family balancing." Many studies were done before to prefer one gender upon another.

When we look at the statistics the chances of having a boy or a girl are almost the same and there is no medical evidence to suggest we can influence

Design: Prospective randomized controlled study.

Setting: The study was conducted at LAMIS Clinic for Obstetrics & Gynecology from January 2015 to February 2022 and still continue up today.

The study started after observation that use of cyclofert female capsules before conceptions leads to more probability to male sex embryos.

Our study contains five thousand patients who received cyclofert female capsules at dose of two capsules per day for at least two months before conceptions. The data was compared with another one thousands patients who receive placebo. The two groups divided into three according to the age group. All patient included were on natural cycles and no ovulation inductions used, irrespective of the parity.

Objectives: To rule out that the observation which is noted that increase male gender embryos related to use of cyclofertfemal capsules or not.

Results: The overall pregnancy rate irrespective of the age was 86.02% (4301/5000) & 66.6% (666/1000) for the cyclofert group and the placebo group respectively. Overall abortion rate for cyclofert group was 1.4% (60/4301) while 9.3% (62/666) for placebo group. 89.69% (3804/4241) of the total pregnancy were male gender in cyclofert Group, while at placebo group it was 41.23 % (249/604), ($p < 0.05$ statistically significant).

Conclusion: Gender selection could be work by non-invasive, not expensive, effective, affordable, and available without complication method by giving cyclofert female capsules, which is, increases the pregnancy rate, reduces the rate of abortion and increases the male gender embryos. Although, there is no guarantee that the baby will actually be one sex or the other, but cyclofertfemal capsules can influence what type of sperm cell will reach an ovum, and fertilize it. Therefore, yes, male gender selection may be technically possible.

Introduction

For decades, gender selection was a problem for many societies, this is because of multiple reasons., the probability of having a boy or a girl depends on whether the sperm that fertilizes the egg is X or Y. Sex selection is an option for couples who want to avoid passing a sex-linked genetic disease to their baby occur with a recessive gene on the X chromosome (Hemophilia and Duchenne muscular dystrophy [1]; Reduce the risk of their baby having a disease more common in certain sexes. For example, a family who has a child with autism spectrum disorder (ASD) might try for a girl because ASD has

a higher male incidence, and their chances of having another affected boy is 25 percent. Alternatively, choose the sex of their baby because of a personal preference "family balancing" [2,3]. Today's sex-selection options aren't equally effective, affordable, or available. The most accurate sex-selection methods are the most expensive and often mean have to undergo invasive fertility treatments (IVF and PGT) [4-10]. This procedure requires to invest significant time and money, and often means have to take fertility drugs before the procedure with potential side effects. There are low-tech, inexpensive gender selection techniques as well. These range from the Shettles and

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Whelan methods to folk wisdom, The American Society for Reproductive Medicine (ASRM) says there's no evidence any of this can influence the sex of the baby. Many countries have taken legislative steps to reduce the incidence of sex-selective abortion. At the International Conference on Population and Development in 1994 over [11,12] 180 states agreed to eliminate "all forms of discrimination against the girl child and the root causes of son preference".

The World Health Organization and UNICEF, along with other United Nations agencies, have found that measures to reduce access to abortion are much less effective at reducing sex-selective abortions than measures to reduce gender inequality. Around 56 million abortions are performed each year in the world, with about 45% done unsafely. India has amongst the worst gender ratios in the world [11,12]. Recent studies have shown that the egg is not passive during the process of fertilization, we say also during sex determination.

Methods

Design

Prospective randomized controlled study.

Setting

The study was conducted at LAMIS clinic for Obstetrics & Gynecology. The period from January 2015 to February 2022 and still continue up today.

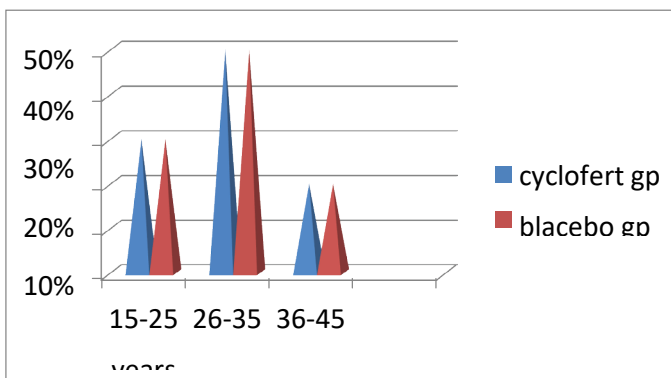
Our study contains five thousand patients who received cyclofert female capsules at dose of two caps per day for at least two months before conceptions. The data were compared with another one thousands patients who receive placebo. The two groups divided into three according to the age group.

The patients undergoing for infertility treatment due to male factor, or assisted reproductive tech either IVF or AIH WHERE excluded, ectopic pregnancy excluded. Patient more than 46 years old were excluded too. All patient included were natural cycles and no ovulation inductions used irrespective of the parity.

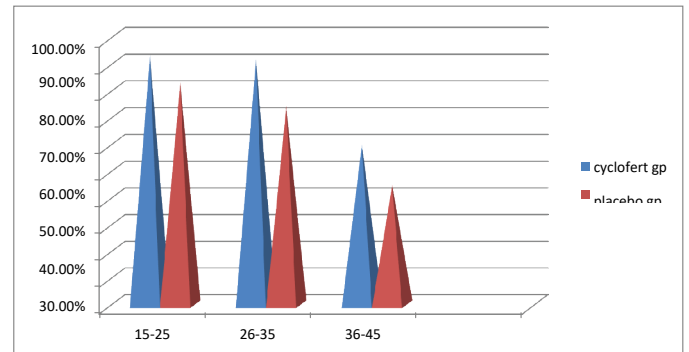
Results

Distribution of both groups according to age

Both cyclofert female group & placebo group are divided to three groups almost the same percentage.



Pregnancy rate during three cycles of both groups



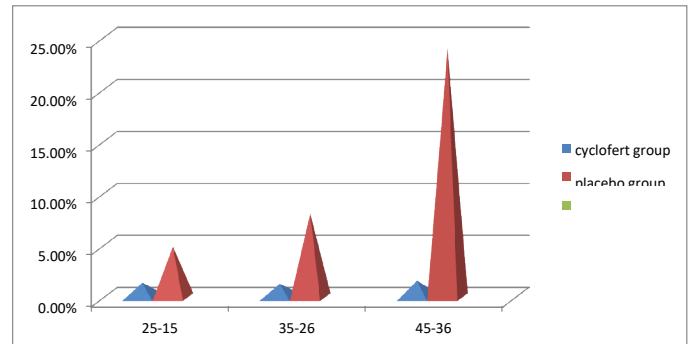
Pregnancy rate at 15-25 age group was (93.3% & 83.3%) at cyclofert group & placebo group respectively.

At 26-35 years age group was (92% & 74.2%) at cyclofert group & placebo group respectively

At 36-45 years age group was (60.2% & 45%) at cyclofert group & placebo group respectively

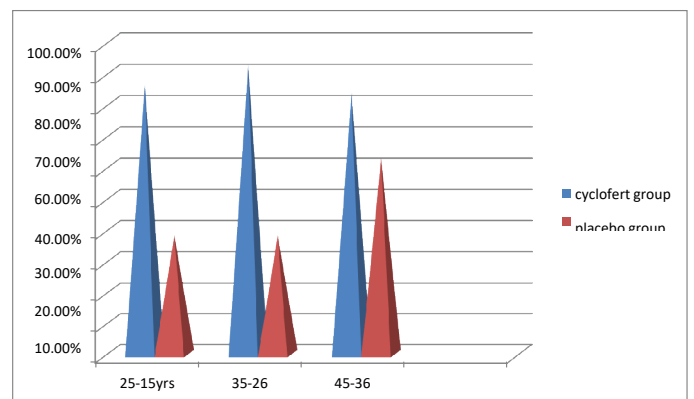
The overall pregnancy rate irrespective of the age was (86.02% & 66.6%) respectively.

Abortion rate at both groups & according to age group



Overall abortion rate at cyclofert group 1.4% while at placebo group 9.3% .(15-25 years age group 20 patients aborted at cyclofert group, while 10 patients aborted at placebo group) (1.4%,4.9%), at 26-35 years group 30 patients aborted at both groups (1.3%, 8.1%). At 36-45 years group 10 patients & 22 patients aborted (1.6% & 24%) respectively.

Gender outcome at each group according to the age group



89.69% (3804/4241) of the total pregnancy were male gender at cyclofert group, while at placebo group were 41.23% (249/604) ($p < 0.05$ statistically significant).

Discussion

We have only touched on sex selection here, and it is a highly complex issue. There are a vast number of reasons people may prefer to have one sex to the other, and there are certain cultures that favor sons over daughters or daughters over sons [3]. Dubuc and Sivia [13] reported that sex selection is a controversial issue. The preference of sons and prenatal sex selection against females have resulted in significant imbalances in sex ratio at birth in several Asian countries, including India and China [11,12]. This issue becomes even more complicated when we consider our ideas of gender and the expectations we may have when choosing the sex of a child.

Many studies done before to prefer one gender upon another, Keep in mind that Mother Nature has already tipped the odds a bit in favor of boys [4-10,14,15,17-19].

According to data from the National Center for Health Statistics, approximately 105 boys are born for every 100 girls. There's always a 50% chance of conceiving a child of the sex wanted [13,19-21].

The probability of having a boy or a girl depends on whether the sperm that fertilizes the egg is X or Y bearing sperm. The origin and maturation of both X and Y spermatozoa are the same, however, certain differences may exist [22-54].

Previous studies proposed a substantial difference between X and Y spermatozoa, however, recent studies (116) suggest negligible or no differences between these spermatozoa with respect to ratio, shape and size, motility and swimming pattern, strength, electric charge, pH, stress response, and aneuploidy. The only difference between X and Y spermatozoa lies in their DNA content [55-59]. Moreover, recent proteomic and genomic studies have identified a set of proteins and genes that are differentially expressed between X and Y spermatozoa. (Therefore, the difference in DNA content might be responsible for the differential expression of certain genes and proteins between these cells [60].

Preimplantation genetic testing (PGT)

A procedure can be done during IVF – after the eggs are fertilized and before an embryo is transferred, one or two cells are removed from an embryo and tested for genetic or chromosomal disorders and/or sex. There are two types of tests: PGT-A and PGT-M are 96

to 97 % accurate at determining the sex of the embryo [61,62] at our study the results show 89.69% of the total pregnancy were male gender at cyclofert group; i.e. almost good result. The PGD procedure is expensive, invasive, can be painful, Fertility drugs can have uncomfortable side effects including: weight gain, bloating, swelling, and blurred vision. Note that some fertility clinics offer preimplantation genetic testing only for medical reasons, and not for sex selection.

Ericsson method

This technique, named for its founder Ronald Ericsson, aims to separate faster-swimming, boy-producing sperm from slower-swimming, girl-producing sperm. It has used in combination with artificial insemination (AI). The technique claims to be 70 to 75% effective [63-69], with more favorable results for conceiving boys than girls. Some independent studies

have supported these claims, while others have contested them. Inexpensive compared to higher-tech methods, noninvasive, relatively safe but there is no guarantee of success. Ericsson has published his own extensive research and claims a success rate of approximately 75 to 80 percent, but evaluations of the test have not been published by other fertility experts or proven independently and AI is not as effective as IVF, and it may take many cycles to achieve a pregnancy, depending on age and fertility.

MicroSort (Flow cytometry)

This technique involves staining the sperm with a fluorescent dye and then separating the male and female sperm cells using a flow cytometer. Once the sperm are identified, the preferred sperm can be placed in the uterus during ovulation using AI or IVF. In its initial clinical trial, MicroSort reported 90% effectiveness for female gender selection and 85 % for male gender selection [70-80]. However, these numbers have not been conclusively confirmed independently. There is some concern that techniques that modify sperm may affect sperm quality. Researchers continue to study this. To use MicroSort, couples must have at least one child and use the technique for the underrepresented gender or be a known carrier of a chromosomal-linked disorder. Note: MicroSort is a trademarked method that is not approved for use in the United States. In 2011, the FDA ordered MicroSort to stop clinical trials and stop using the process; It is still offered in Mexico, Switzerland, North Cyprus, Malaysia, Cambodia, Thailand, and Nigeria.

At-home gender selection techniques

Shettles method: Popular sex selection method named after gynecologist Landrum Shettles. The method is based on a theory of sperm survival. The theory is that sperm bearing a Y chromosome move faster but do not live as long as sperm that carry X chromosomes 75% effective for choosing girls and 80% for choosing boys. In 1970, physician and researcher Landrum.

B. Shettles suggested that couples have intercourse on the day of ovulation or shortly after in order to produce a male offspring. The guidance is involved - even going into detail on who should orgasm first. The method also recommends certain sex positions, claiming that penetration from behind is the best way to conceive a baby boy. Does not require drugs or invasive medical procedure. Shettles explains his methods in a book titled *Your Baby's Sex, Now You Can Choose*. As of 2006, Shettles' book had six revised editions and sold over one million copies [81-85]

Whelan method: Couples should have intercourse two or three days prior to ovulation to increase their chances of conceiving a female infant. To increase the probability of conceiving a male infant, couples should have intercourse between four and six days before ovulation. According to Whelan's research, the method increases the probability of conceiving a male by eighty-six percent and increases the probability of conceiving a female by sixty-six percent. but many experts are doubtful. The Whelan method directly contradicts the Shettles method. The theory here is that the biochemical changes that may favor boy-producing sperm occur earlier in a woman's cycle. The Whelan method was different from the Shettles method. The Shettles method was developed seven years prior to the publication of Whelan's book. Whelan published her method in her book, *Boy or Girl*, in 1977. However, as part of her research, Whelan found that Shettles presented his method for use with natural conception, even though he based his method on studies about

artificial insemination. Whelan cited several studies in her book that demonstrated Shettles' method did not work. Since the publication of Whelan's book, many researchers have refuted her theories regarding the relationship between intercourse timing and an infant's sex. A study published in The New England Journal of Medicine in 1995 refuted all claims that intercourse timing affects the outcome of an infant's sex. That study asserted that there was no association between intercourse timing and sex outcome, There's no guarantee of success [86-90].

Gender selection kits: These at-home kits are based on the Shettles theory. Separate girl and boy kits include a thermometer, ovulation predictor test sticks, vitamins, herbal extracts, and douches that are supposedly intended to favor a specific sex. Kit makers claim a 96% success rate, but some medical experts say the manufacturer's claims have no scientific merit [26]. The success rate claimed by kit makers is questionable, Douching is not recommended, and can actually lead to other problems like vaginal infections [2].

The Chinese gender predictor chart: Legend has it the chart is more than 700 years old and was discovered in a royal tomb near Beijing. The technique involves converting the mother's age and the month of conception to dates on the Chinese lunar calendar, then cross-checking that data on a chart that predicts the baby's sex. The accuracy of the Chinese Gender Calendar Chart is questionable. Some say its accuracy is higher than other gender prediction tools. We've read that about 70% accurate. Others say it's 90% accurate. Researchers at the University of Michigan School of Public Health did a study to test the Chinese lunar calendar method of predicting a baby's sex. They reviewed the records of 2.8 million Swedish births. Then they used a website-customized algorithm to estimate each mother's lunar age and month of conception. When they checked the predictions of the Chinese baby calendar method against the sex of the children who were born, they concluded that the Chinese birth chart was correct about 50 percent of the time -- no more accurate than flipping a coin. There is no scientific basis for the Chinese Gender Calendar Chart, so the most we can guarantee is 50%.

What is the role of cyclofert in sex selection

In addition to the multivitamins and minerals, which contained in a cyclofert capsule needed for good ovum production; there are some natural components. Let's go and see these component and their effects on good ovulation, fertilization, implantation and hence boy sex selection. What factors influence these results, the probability of having a boy or a girl depends on whether the sperm that fertilizes the egg is X or Y bearing, so wondering here, why the cyclofert female capsule taken by the mother can influence whether the sperm that reaches the egg is X or Y, even though it is a random process.

The natural plants component which makes big difference

Licorice dry extract: Licorice is an herb that grows in parts of Europe and Asia. Licorice root contains glycyrrhizin, which can cause side effects when eaten in large amounts. The chemicals in licorice are thought to decrease swelling, decrease cough, and increase the chemicals in the body that heal ulcers. Licorice is used for eczema, swelling of the liver, mouth sores, and many other conditions; it was used traditionally for treating a variety of conditions, including lung, liver, circulatory, and kidney diseases. Today, licorice root is promoted as a dietary supplement for conditions such as digestive problems,

Table. Information about cyclofert female capsules

Composition	Per 1 Capsule
Maritime pine bark dry extract (pinusmaritima)	25 mg
Acetyl-carnitine	25 mg
Co-enzyme Q10	6.250 mg
Zinc	1.875 mg
Vitamin B6	0.75 mg
Vitamin B9 (Folic acid)	0.2 mg
Vitamin B12	0.375 µg
Yam Root dry extract	16 mg
Vitamin E	25 mg
Selenium	7 µg
Omega-3 fish oil TG12/50	225 mg
Licorice dry extract	7 mg

menopausal symptoms, cough, and bacterial and viral infections. Licorice Extract is useful to ease lungs congestion and coughing by helping to loosen thick mucus in airways, which makes cough more productive to expel & so as at the cervical mucus and, B. Shettles using a phase contrast microscope concluded that the small, round headed sperm contained male-producing Y chromosomes, while the large, oval-shaped sperm contained the female-producing X chromosomes [28]. Y sperm: are faster but survive for less time in the female genital tract [91]. X sperm: are slower, but they are more resistant and therefore survive longer. The mucolytic effect of Licorice dry extract facilitate the sperm movements. At the other hand, some researchers have found no morphological differences between human X sperm and Y sperm genotypes; and Y-sperm do not swim faster than X-sperm [92-98]. Scherker [3] reported that Y spermatozoa had a greater tolerance to alkaline pH than X spermatozoa and they moved faster in this media, whereas the reverse being true for acidic pH. To the best of our knowledge, this study among the recent studies that have discovered the role of vaginal pH in fetal sex prediction. It found that there was significant higher rate of male fetuses who were conceived by mothers with alkaline vaginal pH and also there was significant higher rate of female fetuses who were conceived by mothers with acidic vaginal pH. Recently, the study by Oyeyipo et al. [93] aimed to separate X-chromosome and Y-chromosome-bearing spermatozoa using methods based on the viability difference between the X-chromosome and Y-chromosome-bearing spermatozoa, The vaginal environment is generally acidic, while the cervix and uterus are generally alkaline. Licorice dry extract is intended to change the gastric medium to make it more alkaline. Hence, Licorice dry extract change the vaginal environment to make it more alkaline, favor y-bearing sperm to fertilize the egg. (For a boy, the woman's vaginal pH should be 7.5 to 9 [99-104].

Wild yam: Wild yam is a plant that has been promoted as natural DHEA because it contains diosgenin, which can be used in the lab to create estrogen and DHEA. People most commonly use wild yam as a "natural alternative" to estrogen therapy for symptoms of menopause, infertility, menstrual problems, and many other conditions [75,104] is believed to influence hormone balances in a way that can alleviate conditions like morning sickness, premenstrual syndrome, hot flashes, menstrual cramps, vaginal dryness, low libido, and osteoporosis. Herbal

supplements manufacturers will often describe wild yam as "natural estrogen" or "natural DHEA" .used for the commercial synthesis of cortisone, pregnenolone, progesterone, and other steroid products [105].

Capacitated sperm are attracted to progesterone, which is secreted from the cumulus cells surrounding the oocyte. [93,106-110] Wild yam, contains diosgenin needed for Synthesis of cortisone & required for progesterone synthesis. Progesterone binds to the CatSper receptor on the sperm membrane and increases intracellular calcium levels, causing hyperactive motility. The sperm will continue to swim towards higher concentrations of progesterone, effectively guiding it to the oocyte. (chemo attractants that activate and guide sperm to the oocyte [111].

Pine bark extract: They're evergreen trees that grow abundantly in many parts of the northern hemisphere and in some parts of the southern hemisphere as well. In the past, many cultures have used the bark, needles, resin, and nuts of pine trees as medicine [112]. In the 1940s, scientist Jacques Masquelier began studying the health effects of pine bark after learning that indigenous peoples of North America were using pine bark tea to heal scurvy and wounds [133]. It's thought that the extract's antioxidant, antimicrobial, and anti-inflammatory properties have the potential to improve conditions like cancer, heart disease, and neurodegenerative conditions like Alzheimer's disease [112]. Pine bark extract is particularly rich in plant compounds called polyphenols, which are likely responsible for its health-promoting benefits. its constituent antioxidative phenolics are potent neuroprotective agents that can maintain cell viability under oxidative stress [114].

The uterus aids in the steps of capacitation by secreting sterol binding albumin, lipoproteins, and protolytic and glycosidase enzymes such as heparin, It's thought that the Pine bark extract's antioxidant, antimicrobial, and anti-inflammatory properties increase the uterine secretions [93,106,108-110,115]. The sperm acrosome reaction can be stimulated in vitro by substances a sperm cell may encounter naturally, such as progesterone or follicular fluid, as well as the more commonly used calcium ionophore [116,117]. As the sperm approaches the zonapellucida of the egg, which is necessary for initiating the acrosome reaction, the membrane surrounding the acrosome fuses with the plasma membrane of the sperm's head, exposing the contents of the acrosome. The contents include surface antigens necessary for binding to the egg's cell membrane, and numerous enzymes which are responsible for breaking through the egg's tough coating and allowing fertilization to occur [93,104,107,118]. When the sperm enters the vitelline space, receptors on the sperm head called Izumo1 bind to Juno on the oocyte membrane [91]. Once it is bound, two blocks to polyspermy then occur. After approximately 40 minutes, the other Juno receptors on the oocyte are lost from the membrane, causing it to no longer be fusogenic. The cortical reaction and acrosome reaction are both essential to ensure that only one sperm will fertilize an egg [23].

So the effect of cyclofert capsule in gender selection contains good ovum production, good ovulation, alkaline the vagina pH, thinning of the cervical mucus, increase attractive power of the sperm towards the ovum and anti-oxidants, anti-inflammatory effect.

References

1. Almiñana C, Caballero I, Heath PR, et al. The battle of the sexes

starts in the oviduct: modulation of oviductal transcriptome by X and Y-bearing spermatozoa. BMC Genomics. 2014;15(1):293. Published 2014 May 21.

2. Rai P, Ganguli A, Balachandran S, Gupta R, Neogi SB. Global sex selection techniques for family planning: a narrative review. J Reprod Infant Psychol. 2018;36(5):548-560.
3. Griffin DK, Abruzzo MA, Millie EA, Feingold E, Hassold TJ. Sex ratio in normal and disomic sperm: evidence that the extra chromosome 21 preferentially segregates with the Y chromosome. Am J Hum Genet. 1996;59(5):1108-1113.
4. Ishihara K, Warita K, Tanida T, Sugawara T, Kitagawa H, Hoshi N. Does paternal exposure to 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD) affect the sex ratio of offspring?. J Vet Med Sci. 2007;69(4):347-352.
5. Iwata H, Shiono H, Kon Y, et al. Effects of modification of in vitro fertilization techniques on the sex ratio of the resultant bovine embryos. Anim Reprod Sci. 2008;105(3-4):234-244.
6. James WH, Rostron J. Parental age, parity and sex ratio in births in England and Wales, 1968-77. J Biosoc Sci. 1985;17(1):47-56.
7. James WH. The human sex ratio. Part 1: A review of the literature. Hum Biol. 1987;59(5):721-752.
8. Johnson LA. Sexing mammalian sperm for production of offspring: the state-of-the-art. Anim Reprod Sci. 2000;60-61:93-107.
9. Johnson LA, Flook JP, Hawk HW. Sex preselection in rabbits: live births from X and Y sperm separated by DNA and cell sorting. Biol Reprod. 1989;41(2):199-203.
10. Johnson LA, Welch GR, Keyvanfar K, Dorfmann A, Fugger EF, Schulman JD. Gender preselection in humans? Flow cytometric separation of X and Y spermatozoa for the prevention of X-linked diseases. Hum Reprod. 1993;8(10):1733-1739.
11. UN, Population and Development: Programme of Action Adopted at the International Conference on Population and Development, Cairo, Sept. 5–13, 1994, New York: Department for Economic and Social Information and Policy Analysis, UN, 1995.
12. United Nations (UN), Declaration of the Fourth World Conference on Women, Beijing, September 4–15, 1995, New York: UN, 1995.
13. Dubuc S, Sivia DS. Is sex ratio at birth an appropriate measure of prenatal sex selection? Findings of a theoretical model and its application to India. BMJ Glob Health. 2018;3(4):e000675.
14. Flaherty SP, Matthews CD. Application of modern molecular techniques to evaluate sperm sex selection methods. Mol Hum Reprod. 1996;2(12):937-942.
15. Hendriksen PJ, Tieman M, Van der Lende T, Johnson LA. Binding of anti-H-Y monoclonal antibodies to separated X and Y chromosome-bearing porcine and bovine sperm. Mol Reprod Dev. 1993;35(2):189-196.
16. Ruder A. Paternal-age and birth-order effect on the human secondary sex ratio. Am J Hum Genet. 1985;37(2):362-372.
17. Ryan JJ, Amirova Z, Carrier G. Sex ratios of children of Russian pesticide producers exposed to dioxin. Environ Health Perspect. 2002;110(11):A699-A701.
18. van Larebeke NA, Sasco AJ, Brophy JT, Keith MM, Gilbertson M, Watterson A. Sex ratio changes as sentinel health events of endocrine disruption. Int J Occup Environ Health. 2008;14(2):138-143.
19. Curtsinger JW, Ito R, Hiraizumi Y. A two-generation study of

- human sex-ratio variation. *Am J Hum Genet.* 1983;35(5):951-961.
20. Erickson JD. The secondary sex ratio in the United States 1969-71: association with race, parental ages, birth order, paternal education and legitimacy. *Ann Hum Genet.* 1976;40(2):205-212.
 21. Gellatly C. Trends in population sex ratios may be explained by changes in the frequencies of polymorphic alleles of a sex ratio gene. *Evol. Biol.* 2009;36:190-200.
 22. Bean B. Progenitive sex ratio among functioning sperm cells. *Am J Hum Genet.* 1990;47(2):351-353.
 23. Carvalho JO, Silva LP, Sartori R, Dode MA. Nanoscale differences in the shape and size of X and Y chromosome-bearing bovine sperm heads assessed by atomic force microscopy. *PLoS One.* 2013;8(3):e59387.
 24. Chandler JE, Steinholt-Chenevert HC, Adkinson RW, Moser EB. Sex ratio variation between ejaculates within sire evaluated by polymerase chain reaction, calving, and farrowing records. *J Dairy Sci.* 1998;81(7):1855-1867.
 25. Chaudhary I, Jain M, Halder A. Sperm sex ratio (X: Y ratio) and its variations. *Austin J Reprod Med. Infertil.* 2014;1:7.
 26. Chen X, Yue Y, He Y, et al. Identification and characterization of genes differentially expressed in X and Y sperm using suppression subtractive hybridization and cDNA microarray. *Mol Reprod Dev.* 2014;81(10):908-917.
 27. Chen X, Zhu H, Wu C, et al. Identification of differentially expressed proteins between bull X and Y spermatozoa. *J Proteomics.* 2012;77:59-67.
 28. Cui KH. Size differences between human X and Y spermatozoa and prefertilization diagnosis. *Mol Hum Reprod.* 1997;3(1):61-67.
 29. Cui KH, Matthews CD. X larger than Y. *Nature.* 1993;366(6451):117-118.
 30. Eisenberg ML, Murthy L, Hwang K, Lamb DJ, Lipshultz LI. Sperm counts and sperm sex ratio in male infertility patients. *Asian J Androl.* 2012;14(5):683-686.
 31. Geraedts JP. X spermatozoa larger than Y in 1973. *Mol Hum Reprod.* 1997;3(6):545-546.
 32. Goldman AS, Fomina Z, Knights PA, Hill CJ, Walker AP, Hultén MA. Analysis of the primary sex ratio, sex chromosome aneuploidy and diploidy in human sperm using dual-colour fluorescence in situ hybridisation. *Eur J Hum Genet.* 1993;1(4):325-334.
 33. Grant VJ. Entrenched misinformation about X and Y sperm. *BMJ.* 2006;332(7546):916.
 34. Griffin DK, Abruzzo MA, Millie EA, Feingold E, Hassold TJ. Sex ratio in normal and disomic sperm: evidence that the extra chromosome 21 preferentially segregates with the Y chromosome. *Am J Hum Genet.* 1996;59(5):1108-1113.
 35. Hendriksen PJ. Do X and Y spermatozoa differ in proteins?. *Theriogenology.* 1999;52(8):1295-1307.
 36. Hoppe PC, Koo GC. Reacting mouse sperm with monoclonal H-Y antibodies does not influence sex ratio of eggs fertilized in vitro. *J Reprod Immunol.* 1984;6(1):1-9.
 37. Hossain AM, Barik S, Kulkarni PM. Lack of significant morphological differences between human X and Y spermatozoa and their precursor cells (spermatids) exposed to different prehybridization treatments. *J Androl.* 2001;22(1):119-123.
 38. Hu YC, Namekawa SH. Functional significance of the sex chromosomes during spermatogenesis. *Reproduction.* 2015;149(6):R265-R277.
 39. Irving J, Bittles A, Peverall J, Murch A, Matson P. The ratio of X- and Y-bearing sperm in ejaculates of men with three or more children of the same sex. *J Assist Reprod Genet.* 1999;16(9):492-494.
 40. Kaneko S, Oshio S, Kobayashi T, Iizuka R, Mohri H. Human X- and Y-bearing sperm differ in cell surface sialic acid content. *Biochem Biophys Res Commun.* 1984;124(3):950-955.
 41. Kruger AN, Brogley MA, Huizinga JL, et al. A Neofunctionalized X-Linked Ampliconic Gene Family Is Essential for Male Fertility and Equal Sex Ratio in Mice. *Curr Biol.* 2019;29(21):3699-3706.e5.
 42. van DUIJN C Jr. Nuclear structure of human spermatozoa. *Nature.* 1960;188:916-918.
 43. Lobel SM, Pomponio RJ, Mutter GL. The sex ratio of normal and manipulated human sperm quantitated by the polymerase chain reaction. *Fertil Steril.* 1993;59(2):387-392.
 44. Martin RH, Balkan W, Burns K, Rademaker AW, Lin CC, Rudd NL. The chromosome constitution of 1000 human spermatozoa. *Hum Genet.* 1983;63(4):305-309.
 45. Martin RH, Spriggs E, Ko E, Rademaker AW. The relationship between paternal age, sex ratios, and aneuploidy frequencies in human sperm, as assessed by multicolor FISH. *Am J Hum Genet.* 1995;57(6):1395-1399.
 46. McAuliffe ME, Williams PL, Korrick SA, Altshul LM, Perry MJ. Environmental exposure to polychlorinated biphenyls and p,p'-DDE and sperm sex-chromosome disomy. *Environ Health Perspect.* 2012;120(4):535-540.
 47. McDonald E, Watterson A, Tyler AN, McArthur J, Scott EM. Multi-factorial influences on sex ratio: a spatio-temporal investigation of endocrine disruptor pollution and neighborhood stress. *Int J Occup Environ Health.* 2014;20(3):235-246.
 48. Mocarelli P, Gerthoux PM, Ferrari E, et al. Paternal concentrations of dioxin and sex ratio of offspring. *Lancet.* 2000;355(9218):1858-1863.
 49. Perry MJ. Effects of environmental and occupational pesticide exposure on human sperm: a systematic review. *Hum Reprod Update.* 2008;14(3):233-242.
 50. Quinlivan WL, Preciado K, Long TL, Sullivan H. Separation of human X and Y spermatozoa by albumin gradients and Sephadex chromatography. *Fertil Steril.* 1982;37(1):104-107.
 51. Quinlivan WL, Sullivan H. The ratios and separation of X and Y spermatozoa in human semen. *Fertil Steril.* 1974;25(4):315-318.
 52. Scott C, de Souza FF, Aristizabal VHV, et al. Proteomic profile of sex-sorted bull sperm evaluated by SWATH-MS analysis. *Anim Reprod Sci.* 2018;198:121-128.
 53. Shannon M, Handel MA. Expression of the Hprt gene during spermatogenesis: implications for sex-chromosome inactivation. *Biol Reprod.* 1993;49(4):770-778.
 54. Song WH, Mohamed EA, Pang WK, et al. Effect of endocrine disruptors on the ratio of X and Y chromosome-bearing live spermatozoa. *Reprod Toxicol.* 2018;82:10-17.
 55. Umehara T, Tsujita N, Shimada M. Activation of Toll-like receptor 7/8 encoded by the X chromosome alters sperm motility and provides a novel simple technology for sexing sperm. *PLoS Biol.* 2019;17(8):e3000398. Published 2019 Aug 13.
 56. van Munster EB, Stap J, Hoebe RA, te Meerman GJ, Aten JA. Difference in volume of X- and Y-chromosome-bearing bovine

- sperm heads matches difference in DNA content. *Cytometry*. 1999;35(2):125-128.
57. You YA, Kwon WS, Saidur Rahman M, Park YJ, Kim YJ, Pang MG. Sex chromosome-dependent differential viability of human spermatozoa during prolonged incubation. *Hum Reprod*. 2017;32(6):1183-1191.
 58. You YA, Mohamed EA, Rahman MS, et al. 2,3,7,8-Tetrachlorodibenzo-p-dioxin can alter the sex ratio of embryos with decreased viability of Y spermatozoa in mice. *Reprod Toxicol*. 2018;77:130-136.
 59. Zavaczki Z, Celik-Ozenci C, Ovari L, et al. Dimensional assessment of X-bearing and Y-bearing haploid and disomic human sperm with the use of fluorescence in situ hybridization and objective morphometry. *Fertil Steril*. 2006;85(1):121-127.
 60. Rahman MS, Pang MG. New Biological Insights on X and Y Chromosome-Bearing Spermatozoa. *Front Cell Dev Biol*. 2020;7:388.
 61. Gharagozloo P, Aitken RJ. The role of sperm oxidative stress in male infertility and the significance of oral antioxidant therapy. *Hum Reprod*. 2011;26(7):1628-1640.
 62. European IVF-Monitoring Consortium (EIM); European Society of Human Reproduction and Embryology (ESHRE), Kupka MS, et al. Assisted reproductive technology in Europe, 2011: results generated from European registers by ESHRE. *Hum Reprod*. 2016;31(2):233-248.
 63. Check JH, Shanis BS, Cooper SO, Bollendorf A. Male sex preselection: swim-up technique and insemination of women after ovulation induction. *Arch Androl*. 1989;23(2):165-166.
 64. Dominko T, First NL. Relationship between the maturational state of oocytes at the time of insemination and sex ratio of subsequent early bovine embryos. *Theriogenology*. 1997;47(5):1041-1050.
 65. Ericsson RJ, Langevin CN, Nishino M. Isolation of fractions rich in human Y sperm. *Nature*. 1973;246(5433):421-424.
 66. Evans JM, Douglas TA, Renton JP. An attempt to separate fractions rich in human Y sperm. *Nature*. 1975;253(5490):352-354.
 67. Guerrero R. Type and time of insemination within the menstrual cycle and the human sex ratio at birth. *Stud Fam Plann*. 1975;6(10):367-371.
 68. Han TL, Flaherty SP, Ford JH, Matthews CD. Detection of X- and Y-bearing human spermatozoa after motile sperm isolation by swim-up. *Fertil Steril*. 1993;60(6):1046-1051.
 69. Yan J, Feng HL, Chen ZJ, Hu J, Gao X, Qin Y. Influence of swim-up time on the ratio of X- and Y-bearing spermatozoa. *Eur J Obstet Gynecol Reprod Biol*. 2006;129(2):150-154.
 70. Bibbins PE Jr, Lipshultz LI, Ward JB Jr, Legator MS. Fluorescent body distribution in spermatozoa in the male with exclusively female offspring. *Fertil Steril*. 1988;49(4):670-675.
 71. Blottner S, Bostedt H, Mewes K, Pitra C. Enrichment of bovine X and Y spermatozoa by free-flow electrophoresis. *Zentralbl Veterinarmed A*. 1994;41(6):466-474.
 72. Egozcue J, Blanco J, Vidal F. Chromosome studies in human sperm nuclei using fluorescence in-situ hybridization (FISH). *Hum Reprod Update*. 1997;3(5):441-452.
 73. Engelmann U, Krassnigg F, Schatz H, Schill WB. Separation of human X and Y spermatozoa by free-flow electrophoresis. *Gamete Res*. 1988;19(2):151-160.
 74. Karabinus DS. Flow cytometric sorting of human sperm: MicroSort clinical trial update. *Theriogenology*. 2009;71(1):74-79.
 75. Han TL, Ford JH, Webb GC, Flaherty SP, Correll A, Matthews CD. Simultaneous detection of X- and Y-bearing human sperm by double fluorescence in situ hybridization. *Mol Reprod Dev*. 1993;34(3):308-313.
 76. Hassanane M, Kovacs A, Laurent P, Lindblad K, Gustavsson I. Simultaneous detection of X- and Y-bearing bull spermatozoa by double colour fluorescence in situ hybridization. *Mol Reprod Dev*. 1999;53(4):407-412.
 77. Engelmann U, Krassnigg F, Schatz H, Schill WB. Separation of human X and Y spermatozoa by free-flow electrophoresis. *Gamete Res*. 1988;19(2):151-160.
 78. Lechniak D, Strabel T, Bousquet D, King AW. Sperm pre-incubation prior to insemination affects the sex ratio of bovine embryos produced in vitro. *Reprod Domest Anim*. 2003;38(3):224-227.
 79. van Kooij RJ, van Oost BA. Determination of sex ratio of spermatozoa with a deoxyribonucleic acid-probe and quinacrine staining: a comparison. *Fertil Steril*. 1992;58(2):384-386.
 80. Windsor DP, Evans G, White IG. Sex predetermination by separation of X and Y chromosome-bearing sperm: a review. *Reprod Fertil Dev*. 1993;5(2):155-171.
 81. Rorvik DM, Landrum BS. *Your Baby's Sex: Now You Can Choose*. New York City: Bantam Books, 1971.
 82. Rorvik DM, Landrum BS. *How to Choose the Sex of Your Baby: The Method Best Supported by Scientific Evidence*. New York City: Broadway Books, 2006.
 83. Shettles LB. Nuclear morphology of human spermatozoa. *Nature*. 1960;186:648-649.
 84. Shettles LB. Human spermatozoa shape in relation to sex ratios. *Fertil Steril*. 1961;12:502-508.
 85. Shettles LB. Factors influencing sex ratios. *Int J Gynecol Obstet*. 1970;8:643-647.
 86. Blight A. *The Whelan Method of Sex Selection*. Embryo Project Encyclopedia Available at: <http://embryo.asu.edu/handle/10776/13069>
 87. Wilcox AJ, Weinberg CR, Baird DD. Timing of sexual intercourse in relation to ovulation. Effects on the probability of conception, survival of the pregnancy, and sex of the baby. *N Engl J Med*. 1995;333(23):1517-1521.
 88. Whelan E. *Boy or Girl*. New York City: Pocket Books, 1977.
 89. Whelan, E. *Boy or Girl*. New York City: Pocket Books, 1984.
 90. Whelan E. Human Sex Ratio as a Function of the Timing of Insemination within the Menstrual Cycle. *Social Biol*. 1974;21:379- 84.
 91. Pratt NC, Huck UW, Lisk RD. Offspring sex ratio in hamsters is correlated with vaginal pH at certain times of mating. *Behav Neural Biol*. 1987;48(2):310-316.
 92. Ohno S, Wachtel SS. On the selective elimination of Y-bearing sperm. *Immunogenetics*. 1978;7(1):13-16.
 93. Oyeyipo IP, van der Linde M, du Plessis SS. Environmental Exposure of Sperm Sex-Chromosomes: A Gender Selection Technique. *Toxicol Res*. 2017;33(4):315-323.
 94. Penfold LM, Holt C, Holt WV, Welch GR, Cran DG, Johnson LA. Comparative motility of X and Y chromosome-bearing bovine sperm separated on the basis of DNA content by flow sorting. *Mol Reprod Dev*. 1998;50(3):323-327.

95. Pérez-Crespo M, Pintado B, Gutiérrez-Adán A. Scrotal heat stress effects on sperm viability, sperm DNA integrity, and the offspring sex ratio in mice. *Mol Reprod Dev.* 2008;75(1):40-47.
96. Samura O, Miharu N, He H, Okamoto E, Ohama K. Assessment of sex chromosome ratio and aneuploidy rate in motile spermatozoa selected by three different methods. *Hum Reprod.* 1997;12(11):2437-2442.
97. Sarkar S, Jolly DJ, Friedmann T, Jones OW. Swimming behavior of X and Y human sperm. *Differentiation.* 1984;27(2):120-125.
98. Diasio RB, Glass RH. Effects of pH on the migration of X and Y sperm. *Fertil Steril.* 1971;22(5):303-305.
99. Gaber MA, Saleh SA, Allam NH. Effect of vaginal pH in preconceptional fetal sex determination. *Menoufia Med J* 2020;33:1063-6
100. Hamamah S, Gatti JL. Role of the ionic environment and internal pH on sperm activity. *Hum Reprod.* 1998;13 Suppl 4:20-30.
101. Miyazaki R, Fukuda M, Takeuchi H, Itoh S, Takada M. Flow cytometry to evaluate acrosome-reacted sperm. *Arch Androl.* 1990;25(3):243-251.
102. Depypere HT, Comhaire FH. Herbal preparations for the menopause: beyond isoflavones and black cohosh. *Maturitas.* 2014;77(2):191-194.
103. Aitken RJ. Age, the environment and our reproductive future: bonking baby boomers and the future of sex. *Reproduction.* 2013;147(2):S1-S11. Published 2013 Dec 20.
104. Ikawa M, Inoue N, Benham AM, Okabe M. Fertilization: a sperm's journey to and interaction with the oocyte. *J Clin Invest.* 2010;120(4):984-994.
105. Rahman MS, Kwon WS, Pang MG. Prediction of male fertility using capacitation-associated proteins in spermatozoa. *Mol Reprod Dev.* 2017;84(9):749-759.
106. Salicioni AM, Platt MD, Wertheimer EV, et al. Signalling pathways involved in sperm capacitation. *Soc Reprod Fertil Suppl.* 2007;65:245-259.
107. Visconti PE. Understanding the molecular basis of sperm capacitation through kinase design. *Proc Natl Acad Sci U S A.* 2009;106(3):667-668.
108. Battistone MA, Da Ros VG, Salicioni AM, et al. Functional human sperm capacitation requires both bicarbonate-dependent PKA activation and down-regulation of Ser/Thr phosphatases by Src family kinases. *Mol Hum Reprod.* 2013;19(9):570-580.
109. Carson JD, Jenkins RL, Wilson EM, Howell WM, Moore R. Naturally occurring progesterone in loblolly pine (*Pinus taeda* L.): a major steroid precursor of environmental androgens. *Environ Toxicol Chem.* 2008;27(6):1273-1278.
110. Kim JW, Im S, Jeong HR, et al. Neuroprotective Effects of Korean Red Pine (*Pinus densiflora*) Bark Extract and Its Phenolics. *J Microbiol Biotechnol.* 2018;28(5):679-687.
111. Lechniak D, Strabel T, Bousquet D, King AW. Sperm pre-incubation prior to insemination affects the sex ratio of bovine embryos produced in vitro. *Reprod Domest Anim.* 2003;38(3):224-227.
112. Holt WV, Fazeli A. The oviduct as a complex mediator of mammalian sperm function and selection. *Mol Reprod Dev.* 2010;77(11):934-943.
113. CHANG MC. Fertilizing capacity of spermatozoa deposited into the fallopian tubes. *Nature.* 1951;168(4277):697-698.
114. Zhu J, Barratt CL, Lippes J, Pacey AA, Lenton EA, Cooke ID. Human oviductal fluid prolongs sperm survival. *Fertil Steril.* 1994;61(2):360-366.
115. Gómez-Torres MJ, García EM, Guerrero J, et al. Metabolites involved in cellular communication among human cumulus-oocyte-complex and sperm during in vitro fertilization. *Reprod Biol Endocrinol.* 2015;13:123.